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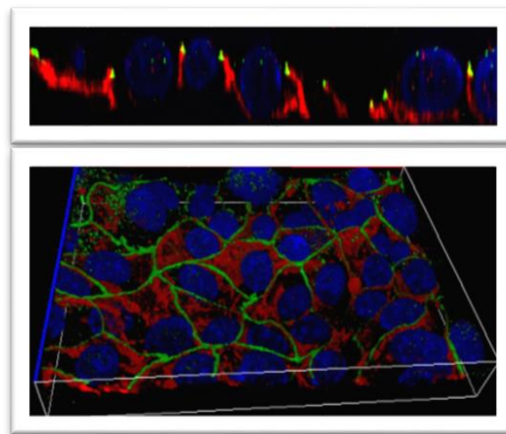
Murine Intestinal Epithelial Cells

CI-muINTEPI



Content

Cell Culture Media & Supplements 2
Getting the cells started after delivery 3
Preparing the Medium for Use and Storage 3
Protocol for cultivating CI-muINTEPI 4
Table 1: Recommended Volumes of Collagen Solution 4
Coating of cell culture plastic ware surfaces 4
Splitting Routine/Maintenance of the cells 5
Table 2: Recommended Volumes of Medium/PBS/TE 5
Freezing/Thawing of the cells 6

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CI-muINTEPI

Cell Culture Media & Supplements

Product Name	***Cat. No.***	***Size***	***Price***
muINTEPI-Medium, *(ready-to-use)* *Consists of basal medium and basal supplements (INS-ME 1005BS)*	INS-ME-1005	500 ml	€ 155.00
Freezing medium	INS-SU-1004	30 ml	€ 29.20
Collagen solution*, (ready-to-use), derived from rat tails, suitable for cell culture*	INS-SU-1017	50 ml	€ 30.00

Recommended for

- CI-muINTEPI
  (Cat.No. INS-CI-1007)
  (CI-muINTEPI-Mx2Luc-10)

Product Description

InSCREENeX muINTEPI Medium is developed for the in vitro cultivation of immortalized murine intestinal epithelial cells (Cat.No. INS-CI-1007).

Quality Control

The medium is subjected to quality control tests and checked for growth characteristics of the immortalized murine intestinal epithelial cell line CI-muINTEPI. In addition, all media have been tested for the absence of microbial contaminants like fungi, bacteria or mycoplasma.

Intended Use

The products are for in vitro use only and not for diagnostic or therapeutic procedures.

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Preparing the Medium for Use and Storage

- The medium and supplements are delivered cooled (@ 2 to 8°C).
- Store the medium @ 4 to 8°C in the dark.
- The supplements should be frozen @ -20°C after arrival.
- It is recommended to aliquot the supplements before and keep them frozen @ -20°C until use.
- If stored properly, the products are stable until the expiry date.
- To give the cultivation media thaw the supplements at 15°C to 25°C and apply the solution to the basal medium.

Getting the cells started after delivery

- The CI-muINTEPI line is being delivered on dry ice. We recommend to thaw the cells on a T25 flask. After two days the medium should be changed.
- As soon as the cells reached adequate confluency (usually 4-5 days after thawing), the cells should be splitted (see protocol at page [5]).
- By starting the culture, the splitting ratio should not exceed 1 : 2.
- In the course of cultivation (after ~2 weeks) the splitting ratio can be extended to 1 : 4 or 1 : 5.

Protocol for cultivating CI-muINTEPI

Coating of cell culture plastic ware surfaces

Material

- Collagen solution (ready-to-use) (Cat.No. INS-SU-1017)
- PBS
- Cell culture plastic ware

Preparation

- Cover the cell culture dish with the Collagen solution (see table 1 below for the required volume).
- Incubate the cell culture dish for at least 120 min (up to overnight) at 37°C in the incubator.
- Aspirate Collagen Solution.
- Wash the coated cell culture dish once with PBS.
- Add cells and media to the coated plate shortly after aspiration of the Collagen Solution.

Plastic ware	***Area in cm²***	***Collagen Solution***
T75	75	2,5 ml
T25	25	1 ml
6 well	9	0,5 ml
12 well	4	0,25 ml
24 well	2	0,1 ml
96 well	0,32	0,05 ml

Table 1: *Recommended Volumes of Collagen Solution*

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Splitting Routine/Maintenance of the cells

Material

- muINTEPI Culture medium (Cat.No. INS-ME-1005)
- PBS
- Cell culture plastic ware
- Trypsin/EDTA solution (TE)
- Sterile Pasteur Pipettes

Preparation

- Check the status of the cells microscopically
- If the cells are 80-90 % confluent, split the cells

Splitting (see ***Table 2*** for corresponding volumes of Medium/PBS/TE)

- Aspirate the cultivation media of the cells with a sterile Pasteur Pipette.
- Wash the cells once with PBS.
- Aspirate the PBS.
- Add TE to the cells.
- Incubate the cells with TE at room temperature or at 37°C until the cells start to detach (check microscopically).
- Resuspend the cells with medium or PBS.
- Transfer an aliquot of the cell solution to a new cell culture dish/flask.
- Add cultivation medium to the cells.
- Cultivate the cells at 37°C.

Plastic ware	*Area in cm²*	*Culture Medium*	*PBS*	*TE*
T75	75	8-10 ml	8-10 ml	2 ml
T25	25	4-5 ml	4-5 ml	1 ml
6 well	9	1,5-2 ml	1,5-2 ml	0,5 ml
12 well	4	1 ml	1 ml	0,2 ml
24 well	2	0,5 ml	0,5 ml	0,2 ml
96 well	0,32	0,1 ml	0,1 ml	0,05 ml

Table 2: Recommended Volumes of Medium/PBS/TE

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Freezing/Thawing of the cells

Material

- Freezing medium (Cat.No. INS-SU-1004)
- PBS
- FBS
- Trypsin/EDTA (TE)
- 15 ml plastic tube
- Vials suitable for freezing in liquid nitrogen

Preparation

Freezing

- Grow the cells to 90 % confluence.
- Wash the cells with PBS.
- Trypsinize the cells with TE.
- Resuspend the cells with 5 ml of PBS, containing 2% FBS.
- Transfer cell suspension in 15 ml plastic tube.
- Spin-down cells @200g for 5 min.
- Aspirate supernatant.
- Resuspend cell pellet with freezing medium (cell concentration $1x10^6$ cells per ml).
- Transfer cell suspension in “freezing” vial.
- Place vials into Mr. Frosty or comparable devices (to slowly cool down vial).
- Place Mr. Frosty in -70°C overnight.
- After 24 h transfer vial from -70°C to liquid nitrogen tank for long term storage.

Thawing

- Pipette 4 ml medium in a 15 ml plastic tube.
- Quickly thaw vial in preheated water bath (@37°C).
- Transfer thawed cell suspension to 15 ml plastic tube containing 4 ml medium.
- Spin-down cells @200g for 5 min.
- Aspirate supernatant.
- Resuspend cell pellet with cultivation medium.
- Transfer cells in desired cultivation device (recommended is T25 flask or 6 well).

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